

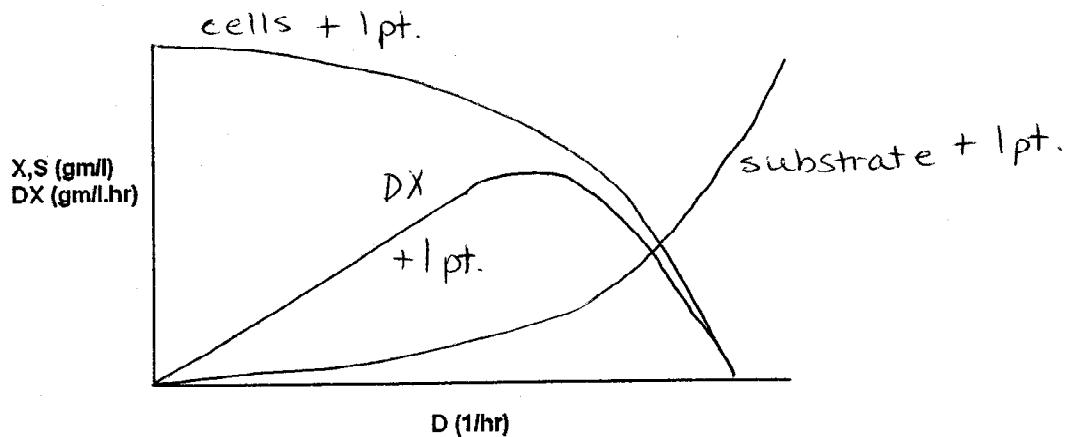
Name: Key

Chemical Engineering 170A

November 9, 2007

Midterm Exam II (100 pts)
Closed Book and Closed Notes
Two 8.5 x 11 in. pages of notes (front and back) allowed

1. Sketch the steady state biomass and substrate concentration (X , S in gm/l) and productivity (DX in gm/l·hr) in a chemostat as a function of dilution rate (D in hr⁻¹) on the plot below. Make sure to label each curve. (3 pts)



2. Show that $\mu = D$ for a chemostat at steady state with a sterile feed (4 pts).

2 pts writing mass balance

$$\frac{VdX}{dt} = F(X_0 - X) + \mu X V$$

mass balance biomass

$$\text{at s.s. } 0 = D(X_0 - X) + \mu X$$

sterile feed $X_0 = 0$ 2 pts. final

$$\mu X - DX = 0 ; (\mu - D) X_{ss} = 0 \quad X_{ss} \neq 0 \text{ so answer}$$
$$\mu - D = 0 \rightarrow \mu = D$$

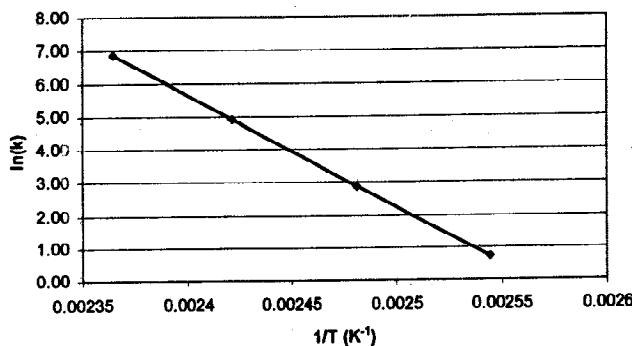
3. You know the power input (in Watts) to a six-blade pitched-blade impeller ($W/D = 1/8$) used in a continuously stirred tank bioreactor. Assume you know the diameter of the impeller and the impeller speed. The density of the fermentation broth can be taken as that of water. Briefly explain, in words, how you would calculate μ_{app} , the apparent viscosity of the fermentation broth (6 pts). 2 pts. power #

Calculate the power # with the diameter D and the

Speed N of the impeller and ρ the density of the broth.

Use chart in Fig 5.20 to find the Re number for the calculated power #. Use Curve 4 for a 6-blade, pitched-blade impeller. From knowing the Re # and D, N, ρ , you can calculate μ_{app} , the apparent viscosity of the broth. 2 pts. for mentioning Re #

4. The following plot relates the rate constant for cell death (k , min^{-1}) of the organism *B. stearothermophilus* to temperature (deg K).



- a) What is the activation energy, E_a , for thermal cell death (6 pts)?

$$\text{Slope} = -E_a/R \quad \ln k = \ln k_0 - \frac{E_a}{R} \frac{1}{T}$$

$$\text{Slope} = \frac{0.80 - 6.90}{0.00254 - 0.00237} = -35882 \text{ J/K} \quad 3 \text{ pts. calculation}$$

$$-35882.4 \text{ K} = -E_a/R \quad E_a = R(35882.4 \text{ K}) \quad 3 \text{ pts. final answer}$$

$$E_a = \frac{8.314 \text{ J}}{\text{K mole}} \times 35882.4 \text{ K} = \boxed{298.3 \text{ kJ/mole}}$$

5. The enzyme urease is immobilized on non-porous polymer beads and is used in a plug flow reactor to convert urea to CO_2 and ammonia. Assume the flow rate is high enough to overcome any external mass transfer effects. If the initial urea concentration is 25 mM, what residence time is needed to convert 80% of the urea to CO_2 and ammonia (6 pts)?

Useful parameters:

$$S_A = 0.80$$

$$\text{Urease } V_{\max} = 10 \text{ mmoles/min} \cdot \text{mole}$$

$$K_M = 20 \text{ mM}$$

$$\hat{T} = \frac{C_{AO} S_A}{V_{\max}} - \frac{K_M}{V_{\max}} \ln(1 - S_A) \quad 3 \text{ pts. equation}$$

$$\hat{T} = \frac{25 \text{ mMoles}}{k} \cdot \frac{\text{min} \cdot k}{10 \text{ mMoles}} \cdot 0.8 - \frac{20 \text{ mMoles}}{k} \cdot \frac{\text{min} \cdot k}{10 \text{ mMoles}} \cdot \ln(0.2)$$

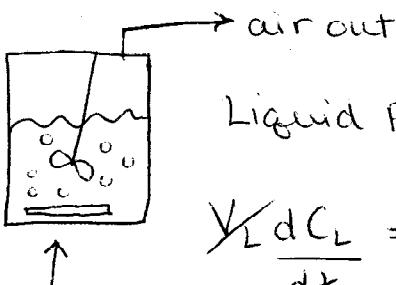
3 pts. calculation and answer

$$\boxed{\hat{T} = 5.22 \text{ mins}}$$

6. E. coli have a maximum respiration rate, $qO_{2\max}$, of 240 mg O₂/gdw·hr. It is desired to achieve a cell mass of 20 gdw/l. The $k_L a$ is 120 h⁻¹ in a 1000 l reactor (800 l liquid volume). A gas stream enriched in oxygen is bubbled through a batch bioreactor to give a value of $C_L^* = 28$ mg/l. If oxygen becomes growth limiting, growth and respiration are slow and qO_2 can be written as

$$qO_2 = \frac{qO_{2\max} C_L}{0.2 \frac{\text{mg}}{\text{l}} + C_L}$$

where C_L is the dissolved oxygen concentration in the fermentor. What is C_L when the cell mass is at 20 g/l (25 pts)?



Liquid Phase O₂ Balance 5 pts. liquid phase mass

$$\cancel{\sum} \frac{dC_L}{dt} = k_L a (C_L^* - C_L) \cancel{\sum} - qO_2 X \cancel{\sum} \text{ balance}$$

In a fed-batch system if oxygen transport is growth limiting then the rate of consumption of oxygen by the cells equals transport of oxygen into the liquid and $\frac{dC_L}{dt} = 0$

pseudo steady-state $0 = k_L a (C_L^* - C_L) = qO_2 X$ 5 pts. for equating

cells aren't growing $R_L a (C_L^* - C_L) = \frac{qO_{2\max} C_L}{0.2 \frac{\text{mg}}{\text{l}} + C_L} X$ $R_L a = 120 \text{ h}^{-1}$ the two

$$120 (28 - C_L) = \frac{240 C_L (20)}{0.2 + C_L} \quad qO_{2\max} = 240 \frac{\text{mg O}_2}{\text{g} \cdot \text{hr}}$$

5 pts. for putting in #'s

$$3360 - 120 C_L = \frac{4800 C_L}{0.2 + C_L}; \quad 4800 C_L = (0.2 + C_L)(3360 - 120 C_L)$$

$$4800 C_L = 672 - 24 C_L + 3360 C_L - 120 C_L^2 \quad \begin{matrix} 5 \text{ pts. for} \\ \text{solving} \end{matrix}$$

$$120 C_L^2 + 1464 C_L - 672 = 0 \quad a = 1 \quad b = 12.2 \quad c = -5.6$$

$$C_L^2 + 12.2 C_L - 5.6 = 0 \quad C_L = -12.2 \pm \sqrt{(12.2)^2 - 4(1)(-5.6)}$$

$$C_L = \frac{-12.2 \pm 13.09}{2} \quad \boxed{C_L = 0.45 \text{ mg/l}} \quad \begin{matrix} 3 \\ 5 \text{ pts. final answer} \\ \text{other root gives a } \ominus C_L \end{matrix}$$

2 (1)

7. A continuous sterilizer at 120 °C runs at a flow rate such that the flow is turbulent. For fully developed turbulent flow, the modified Peclet number, ($d_t \cdot u_s / D$) reaches an upper limit of 3.33. The ratio of reactor length to reactor diameter (L/d_t) is 100, and the reactor volume is 5 l. The unsterilized medium contains 10^4 spores/l and the reactor has a residence time of 5 minutes.

- a) Calculate the value of the Pe for this reactor. Can you assume the continuous sterilizer behaves as a plug flow reactor? Why or why not? (10 pts)

$$\text{modified Pe} = 3.33 = \frac{d_t \cdot u_s}{D} \quad Pe = \frac{L \cdot u_s}{D} \quad 5 \text{ pts. for calculating Pe}$$

$$L/d_t = 100$$

$$dt = \frac{L}{100}$$

$$3.33 = \frac{L \cdot u_s}{D \cdot 100}$$

$$\frac{L \cdot u_s}{D} = 3.33 = Pe$$

5 pts.
explanation so we can assume the continuous reactor is a plug flow reactor

- b) The cell death rate constant k is 2.0 min^{-1} at 120°C . What is the probability of contamination for the medium exiting the reactor after one residence time (10 pts)?

5 pts. for equation - $P(t) = \text{probability of contamination}$

$$P(t) = (1 - e^{-k\bar{t}})^{N_0} \quad N_0 = \# \text{ of spores}$$

$$\bar{t} = 5 \text{ min}$$

$$R = 2 \text{ min}^{-1}$$

$$\text{in 1 residence time } N_0 = \frac{10^4 \text{ spores}}{\text{l}} \times 5 \text{ l}$$

$$1 - P(t) = 1 - (1 - e^{-2(5)})^{50,000} \quad 5 \text{ pts. for working out solution & for answer} \quad N_0 = 50,000$$

$$= 0.897 \quad 89.7\% \text{ probability of contamination}$$

- c) What parameters, if any, can you adjust to decrease the probability of contamination (5 pts)?

To decrease probability of contamination you can increase the temperature or increase the residence time (by either increasing the volume of the reactor or decreasing the flow rate).

5 pts. for explanation

(2.5 pts. for temp, 2.5 pts. for residence time)

8. You are working for a biotechnology company in the Bay Area that wants to produce an antibody from chinese hamster ovary (CHO) cells in a perfusion bioreactor with a residence time of 2 hours. Data from a batch culture (started from an inoculum of 10^4 cells/ml show that the net specific growth rate at 10 hours of culture is 0.5 hr^{-1} and the growth of CHO cells during the exponential phase can be described by the following:

$$\frac{dX}{dt} = \mu X$$

- a) If you plan to operate your perfusion bioreactor at a specific growth rate matching that of the batch system at 10 hours, what is the predicted steady-state value for biomass in the perfusion bioreactor (15 pts)?

5 pts. equation $X_{ss}^P = \frac{F}{V} \int_0^t X^B(t) dt$ $X^B(t) = X_0 e^{\mu t} = 10^4 e^{0.5t}$
 $F/V = \frac{1}{2 \text{ hrs}} = 0.5 \text{ hr}^{-1}$

$$X_{ss}^P = \frac{0.5 \cdot 10^4 \text{ cells}}{\text{h mL}} \times \int_0^{10} e^{0.5t} dt \quad 5 \text{ pts. solving integral}$$

$$X_{ss}^P = 5000 \frac{\text{cells}}{\text{mL h}} \times \left[\frac{0.5}{0.5} e^{0.5t} \right]_0^{10} = \frac{5000}{0.5} \left(e^{0.5(10)} - e^0 \right)$$

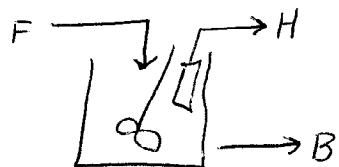
$$X_{ss}^P = 10000 (148.4 - 1) = 1.47 \times 10^6 \text{ cells/mL}$$

5 pts. final answer

- b) The specific antibody production rate in CHO cells is $10 \text{ pg/cell}\cdot\text{hr}$, and is growth associated. Calculate the expected steady-state concentration of antibody in the perfusion bioreactor (10 pts).

mass balance on P

5 pts. mass $\frac{V dP}{dt} = -(H+B)P + \nu X V$



balance (a) s.s. $0 = -\frac{FP}{V} + \nu X \quad H+B=F$

equation $\frac{FP}{V} = \nu X ; P_{ss} = \frac{V}{F} \nu X_{ss}^P$

5 pts. final $P_{ss} = 2 \text{ hr} \times \frac{10 \text{ pg}}{\text{cell}\cdot\text{hr}} \times \frac{1.47 \times 10^6 \text{ cells}}{\text{mL}} = 2.94 \times 10^7 \frac{\text{pg}}{\text{mL}}$

answer calculations $P_{ss} = 29.4 \text{ mg/mL}$